# Reduction of cross contamination and extension of shelf life of ready to cook chicken meat treated with extracts of selected medicinal plants

# By Thikshani Somarathna

Ph.D. 2018

Reduction of cross contamination and extension of shelf life of ready to cook chicken meat treated with extracts of selected medicinal plants

By Thikshani Somarathna



Thesis submitted to the University of Sri Jayewardenepura for the award of the Degree of Doctor of Philosophy

## Thesis Declaration by Author

The work described in this thesis was carried out by me under the supervision of Dr.

NS Weerakkody, Prof. KKDS Ranaweera, Dr. GAS Premakumara Dr. Mayuri

Thammityagodage and a report on this has not been submitted in whole or in part to
any university or any other institution for another Degree \* Diploma\*.

T. Somarathna

(Signature of the candidate)

We certify that the above statement made by the candidate is true and that this thesis is suitable for submission to the University for the purpose of evaluation

Mimsa

Dr. NS Weerakkody

Principle Supervisor

Senior Lecturer.

Department of Agricultural and

Plantation Engineering.

Faculty of Engineering Technology,

The Open University of Sri Lanka

Date 17/05/2019

Prof. KKDS Ranaweera

Internal- Supervisor

Senior Professor,

Department of Food Science and Technology.

Faculty of Applied Science.

University of Sri Jayewardenepura.

Sri Lanka

Date 17/05/2019

External-Supervisor

Director.

Industrial Technology Institute.

363, Buddaloka Mawatha

Colombo 7, Sri Lanka

Date 17/05 /2019

14. h. Chaming excelon

Dr. Mayuri Thanmitiyagodage

External-Supervisor

Head/ Animal Center.

Medical Research Institute.

Borella, Colombo, Sri Lanka

Date 17/05/2019

M. G. Thammitiyegoriage B.v.Sc M On

Voternan Surgabil

Medical Research institution Colombo.

### Certification of the Supervisors

We certify that the the candidate has incorporated all corrections, addition and ammenmendts recommended by the examiners to this version of the PhD thesis.

Minsz

Dr. NS Weerakkody Principle Supervisor Senior Lecturer, Department of Agricultural and Plantation Engineering, Faculty of Engineering Technology, The Open University of Sri Lanka

Date 17/05/2019

Dr. GAS Premakumara External-Supervisor External-Supervisor Director/Senior Research Fellow,

Industrial Technology Institute, 363, Buddaloka Mawatha

Colombo 7, Sri Lanka

Date 17/05/2019

M. h. Themming & len Dr. Mayuri Thanmitiyagodage

Department of Food Science and Technology,

Head/ Animal Center.

Prof. KKDS Ranaweera

Faculty of Applied Science,

University of Sri Jayewardenepura,

Date 17/07/2019

Internal-Supervisor

Senior Professor,

Sri Lanka

Medical Research Institute.

Borella, Colombo, Sri Lanka

Date ... 17/05/2019

M. G. Thammitte and range page Man Vetores - 1,500

Medical Research institution Colombo.

#### **Table of Contents**

	Page No
DECLARATION BY AUTHOR	I
DECLARATION OF THE SUPERVISOR	II
CERTIFICATION OF THE SUPERVISORS	III
TABLE OF CONTENTS	IV
LIST OF TABLES	IX
LIST OF FIGURES	X
ABBREVIATIONS	XII
ACKNOWLEDGEMENTS	XV
ABSTRACT	XVI
CHAPTER 1.0 INTRODUCTION	1
1.1 Objectives	5
1.1.1 Main objective	5
1.1.2 Specific objectives	5
CHAPTER 2.0 LITERATURE REVIEW	7
2.1 Medicinal plants and uses	7
2.2 Natural phytochemicals	8
2.3 Food additives	9
2.3.1Food preservatives	10
2.3.2 Mechanisms of antimicrobial action	11
2.3.3 Factors affecting antimicrobial activity	13
2.4 Medicinal plants used for the study	13
2.4.1 Alpinia malaccensis	14
2.4.2Alpinia purpurata	17
2.4.3Terminalia catappa	20
2.4.4 Curcurma zedoaria	22

2.5 Herb extracts	26
2.6 Solvent extraction methods	28
2.6.1Soxhlet extraction	28
2.6.2 Maceration	29
2.6.3Hydro distillation	30
2.6.4 Ultrasound-assisted extraction (UAE)	30
2.6.5 Supercritical fluid extraction (SFE)	30
2.6.6 Pressurized liquid extraction (PLE)	31
2.7 Food borne pathogens	32
2.7.1 Listeria monocytogenes	32
2.7.2 Staphyloccocus aureus	33
2.7.3 SalmonellaTyphimurium	34
2.7.4 Escherichia coli	35
2.8 Methods to determine antimicrobial activity of plant extracts	36
2.8.1 Agar disk-diffusion method	37
2.8.2 Agar well diffusion method	39
2.8.3Broth dilution method	39
2.8.4 Agar dilution method	41
2.9 Methods of antioxidant determination in a food system	46
2.10 Methods of toxicity determination	50
2.10.1 Cytotoxicity assay	50
2.10.2 Animal test	52
2.11Chicken meat preservation	55
CHAPTER 3.0 MATERIALS AND METHODS	58
3.1 Plant collection and authentication	58
3.2 Chemicals and reagents	58

3.3 Preparation of extracts	59
3.4 Test micro organisms	59
3.5 Disc-diffusion assay	60
3.6 Broth dilution assay	61
3.7 Total phenol content	62
3.8 Chemical composition of the plants	62
3.9 Purification of major chemical compound	63
3.9.1 Thin layer chromatography of plant	extract63
3.9.2 TLC bio-autography	63
3.9.3 Isolation of active chemical compou	nd64
3.10 Nuclear Magnetic Resonance Spectroscopy	64
3.11 Synergistic antimicrobial activity	65
3.11.1 Broth dilution method	65
3.11.2 Agar well diffusion method	65
3.12 Anti-biofilm activity of A. malaccensis	66
3.12.1 Inhibition of initial cell attachment	66
3.12.2 Inhibition of biofilm formation and	l development67
3.12.3 Inhibition biofilm formation	67
3.13 Scanning Electron Microscopy	68
3.14 Synergistic anti-biofilm activity of A. malaccen	sis and T. catappa69
3.15 Active toxicity study using animals	
3.15.2 Acute oral toxicity study	70
3.15.3 Histopathology study	71
3.16 Determination of mammalian toxicity using al	ternative animal studies71
3.16.1 Cell culture	71

	3.16.2 MTT assay for assessment of cell viability	72
3.17 I	dentification of morphological changes	73
	3.17.1 Assessment of cell death AO/EB fluorescentassay	73
	3.17.2 Assessment of nuclear features using Hoechst 33528 staining	ıg73
	3.17.3 Comet assay for genotoxicity assessment	74
3.18 I	Effect of plant extract combination on food matrix	75
	3.18.1 Preparation and inoculation of chicken meat	75
	3.18.2 Marination of chicken meat	76
	3.18.3 Microbial Enumeration	76
	3.18.4 Determination of thiobarbituric acid reactive substances	77
	3.18.5 pH measurements	78
	3.18.6 Color measurements	78
3.19	Statistical Analysis	79
CHAPTER	R 4.0 RESULTS	79
4.1 I	Disk diffusion assay	79
4.2 B	Broth dilution assay	81
4.3 T	Total phenol content	83
4.4 (	Gas chromatography mass spectrometry analysis	83
4.5 T	TLC bioautography	85
4.6 P	Purification of 1' Aacetoxy Chavicoal Acetate	86
4.7 A	Antimicrobial activity of pure compound	87
4.8 N	Nuclear Magnetic resonance	88
4.9 S	Synergistic antimicrobial activity	89
	4.9.1 Broth dilution method	89
	4.9.2 Synergistic antimicrobial activity using agar-well diffusion n	nethod
4.10	Single and synergistic anti-biofilm activity	92

4.11 A	anti-biofilm activity of A. malaccensis by scanning electron micr	oscopy
4.12 A	cute oral toxicity studies using animals	96
	4.12.1 Effect on body weights	97
	4.12.2 Effect on feed and water intake	98
	4.12.3 Effect of extract on the weight of body organs	101
	4.12.4 Biochemical response of rats	102
	4.12.5 Hematological responses of rats	102
	4.12.6 Histopathological observation of organs	103
4.13 T	Γoxicity studies using alternative methods (Cyto-toxicity)	107
	4.13.1 Cytotoxic activity of crude extracts of A. malaccensis	107
	4.13.2 Cytotoxic activity of crude extracts of <i>T. catappa</i>	109
	4.13.3 Microscopic feature of AO/EB staining	111
	4.13.4 Microscopic features of nucleus adopting Hoechst staining	g 113
	4.13.5 Identification of DNA damage using comet assay	114
<b>4.14</b> E	Effect of plant extract combinations on food matrix	115
	4.14.1 Effect of combination of plant extract on <i>L. monocytogene</i>	es on
	chicken samples stored at 4 °C	115
	4.14.2 Lipid peroxidation in chicken stored at 4 °C	116
	4.14.3 Effect of plant mixtures on S. aureus on chicken samplesa	t 8°C
	4.14.4 Lipid peroxidation in chicken at 8 °C	118
	4.14.5 pH of chicken stored at 4 °C and 8 °C	119
	4.14.6 Color changes of chicken stored at 4 °C and 8 °C	119
CHAPTER 5	5.0 DISCUSSION	121
CHAPTER 6	6.0 CONCLUSIONS AND RECOMMENDATIONS	140
REFERENC	CE	147
APPENDIX.		164

Table 1. Antibacterial activity of the medicinal plant tested for the study
Table 2. Total phenol and Antioxidant activity assay methods
Table 3. Disk diffusion assay (diameter of inhibition zone) of underutilized plant
extracts
Table 4. The MIC and MBC values (mg/ml) of underutilized plant extracts against food-
borne pathogens
Table 5. Total phenol content of different solvent extract of the plant83
Table 6. GC-MS results in ethanol and hexane extractof A. malaccensis
Table 7. GC-MS data of <i>T. catappa</i> ethanol extracts
Table 8.Synergistic antibacterial activity of A. malaccensis and T. catappa90
Table 9. Synergistic antibacterial activity using agar well diffusion assay92
Table 10. The effect of plant extract on biofilm-adhesion, growth and formation94
Table 11. Mortality of Wistar rats administered crude extract A. malaccensis and T.
catappa96
Table 12. Organ weights (g) of the female rats
Table 13. Serum biochemical parameters of female rats
Table 14. Serum hematological values of female rats
Table 15. Inoculated $L$ monocytogenes and aerobic plate count on chicken meat at 4 $^{0}\mathrm{C}$
storage
Table 16. TBAR values for chicken stored at 4°C
Table 17. Inoculated S. aureus and aerobic plate counts on chicken treated samples
stored at 8 °C
Table 18. TBAR value for the chicken stored in 8°C
Table 19.pH of chicken stored at 4°C and 8°C
Table 20. Colour changes of the storage of chicken meat at 4°C
Table 21. Colour changes of the storage of chicken meat at 8 °C

Page No

**List of Tables** 

List of Figures Page No

Figure 1.Different plant parts of A. malaccensis(a) Plant (b) Inflorescence (c) Rhizome
Figure 2.Different plant parts of A. purpuata (a) Leaf (b) Inflorescence (c) Rhizome 17
Figure 3.Different plant parts of <i>T. catappa</i> (a) Leaf (b) Fruit
Figure 4. Cucurma zedoria (a) Leaf (b) Rhizome
Figure 5. GC-MS chromatogram of <i>T. catappa</i> ethanol extract
Figure 6. (a) TLC separations in solvent system IV (b) Bio-autography against S. aureus
11386
Figure 7. GC-MS chromatogram of purified compound of <i>A. malaccensis</i>
Figure 8. Mass spectrum of the isolated compound from the A. malaccensis
Figure 9. (a) DIZ of A. malaccensis pure compound (b) DIZ of crude extract of A
malaccensisagainst S. aureus 113
Figure 10. Structure of the isolated compound (1'ACA)
Figure 11. SEM image of S. aureus 113biofilm on stainless steel surfaces
Figure 12. SEM image of <i>L. monocytogenes</i> biofilm on stainless steel surfaces96
Figure 13. Mean body weight of rats administered with A. malaccensis extract97
Figure 14. Mean daily weight gain of rats administered with <i>T. catappa</i> extract98
Figure 15. Mean daily feed intake of rats administered with A. malaccensis
Figure. 16 Mean daily water intake of rats administered with A. malaccensis extract99
Figure. 17 Mean daily feed intake of rats administered with <i>T. catappa</i> extract 100
Figure 18. Mean daily water intake of rats administered with <i>T. catappa</i> extract 101
Figure 19. Photomicrograph of the liver section of rats (A) Control (B) $A$
$malaccensis~300~\mathrm{mg/ml}(C)~A.~malaccensis~2000~\mathrm{mg/ml}~(D)~T. catappa~300~\mathrm{mg/ml}~(E)~T. catappa~3000~\mathrm{mg/ml}~(E)~T. catappa~300~\mathrm{mg/ml}~(E)~T. catappa~300~mg/ml$
catappa 2000 mg/ml x 20
Figure 20. Photomicrograph of the kidney section of rats (A) Control (B) $A$
malaccensis 300 mg/ml(C) A. malaccensis 2000 mg/ml (D) T.catappa 300 mg/ml (E) T
catappa 2000 mg/ml X 20
Figure 21. Photomicrograph of the heart section of rats (A) Control (B) $A$
malaccensis 300 mg/ml(C) A. malaccensis 2000 mg/ml (D) T.catappa 300 mg/ml (E) T
catappa 2000 mg/ml

Figure 22. Photomicrograph of the lung section of rats (A) Control (B) A.
$malaccensis~300~\mathrm{mg/ml}~(C)~A.~malaccensis~2000~\mathrm{mg/ml}~(D)~T. catappa~300~\mathrm{mg/ml}~(E)~T. catappa~300~mg/ml$
catappa 2000 mg/ml
Figure 23. Photomicrograph of the spleen section of rats (A) Control (B) A.
malaccensis 300 mg/ml(C) A. malaccensis 2000 mg/ml (D) T.catappa 300 mg/ml (E) T.
catappa 2000 mg/ml
Figure 24. Cyto-toxicity of A. malaccensis crude extract against A549 (µg/ml) 107
Figure 25. Cyto-toxicity of A. malaccensis crude extract against HepG2 (µg/ml) 108
Figure 26. Cyto-toxicity of A. malaccensis crude extract against 3T3 (µg/ml) 108
Figure 27. Cyto-toxicity of A. malaccensis crude extract against COS7 (µg/ml) 108
Figure 28. Cyto-toxicity of <i>T. catappa</i> crude extract against A549 (µg/ml) 109
Figure 29. Cyto-toxicity of <i>T. catappa</i> crude extract against HepG2 (µg/ml)110
Figure 30. Cyto-toxicity of <i>T. catappa</i> crude extract against 3T3 (µg/ml)110
Figure 31. Cyto-toxicity of <i>T. catappa</i> crude extract against COS 7 (µg/ml)110
Figure 32. Morphological assessment of apoptosis and necrosis
Figure 33. (A)Morphological features of nuclei observed for control and extract-treated
cells stained with Hoechst 33258 (B) Percentage of cells with normal and abnormal
nuclei
Figure 34. Assessment of DNA damage adopting comet assay

#### **Abbreviaztions**

ABTS 2,2'-azinobis-(3-ethylbenzothiazoline-6-sulfonate)

A549 Lung carcinoma cell line
ACA Acetoxychavicol acetate

AC Acridine orange

ANOVA Analysis of variance

ATCC American type culture collection

BHI Brain heart infusion

BHA Butylated hydroxyanisol
BHT Butylated hydroxytoluene

BPA Baird-Parker agar

CDC Centers for Disese Control and Prevention

CFU Colony forming unit

COS7 Monkey Fibroblast Cell Line

COSY Correlation Spectroscopy
CORL-23 Lung Carcinoma Cell Line

CLSL Clinical Laboratory Standard Institute

DEPT Distortionless enhancement by polarization transfer

DMSO Dimethyl sulfoxide

DMEM Dulbecco's Modified Eagle's medium

DIZ Diameter of inhibition zone

EO Essential oils

EB Ethidium Bromide

EDTA Ethylenediaminetetraacetic acid
ETEC Enterotoxigenic *Escherichia coli* 

ET Electron transfer

ESE Enhance Solvent Extraction

FAO Food and Agricultural Organisation

FBS Fetal Bovine Serum

FCR Folin-Ciocalteus reagent

GAE Gallic acid equivalent

GC-MS Gas chromatography-mass spectrometry

HAT Hydrogen Trasfer
HCA Hydroxycitric acid

HHP High hydrostatic pressure

HMBC Heteronuclear Multiple Bond Correlation

HePG2 Hepatocellular carcinoma cell line
HSPE High Pressure Solvent Extraction

MDA Malondialdehyde

MAP Modified atmosphere packaging

MDR Multiple drug resistance

MH Mueller Hinton

MIC Minimum inhibitory concentration

MLC Minimum lethal concentration

MPN Most probable number

MRSA Methicillin resistant Staphylococcus aureus

MSSA Methicillin sensitive Staphylococcus aureus

MTT 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide

NCCS National Center for Cell Science

NMR Nuclear magnetic resonance spectroscopy

NOEAL No observed Adveced Effect Level

OD Optical density

OECD Organization for Economic Cooperation and Development

ORAC Oxygen Radical Absorbance Capacity

PALCAM Polymyxin Acriflavine Lithium Chloride Ceftazidime

PBS Phosphate Buffered Solution

PC3 Prostate adenocarcinoma cell line

PCA Plate count agar

PLE Pressured Liquied Extraction

PV Peroxide value

ROS Reactive Oxygen Species

RBC Red Blood Count
RTE Ready to Cook

SEM Scanining electron microscopy

TBA Thiobarbituric acid

Thiobarbituric acid reactive substances **TBARS** 

Total phenolic content TPC

TLC Thin layerchromatography

TSA Tryptic soy agar Tryptic soy broth **TSB** VP

Vacuum pack

Xanthine oxidase XOD

XLD Desoxycholate agar

Violet red bile glucose agar **VRBG** 

WBC White blood count

#### Acknowledgments

First and foremost, I would like to thank my principle supervisor, Dr. Nimsha Weerakkody whose insight launched a greater part of my study and the co-supervisors: Prof. KKDS Ranaweera, Dr. GAS Premakumara and Dr. Mayrui Thanmitiyagodage for their valuable guidance, constructive criticism, and encouragements given throughout my study. Their generosity in sharing knowledge and practical skills in different expertise were greatly assisting me to stand as a knowledgeable independent researcher. I wish to thank National Research Council (NRC12-54),Government of Sri Lanka, for the financial support given to carry out the research. I am grateful for being awarded partial scholarship ISRF from Government of India for doing cell culture toxicity studies with the guidance of Prof. MA Akbarsha and Associate Prof. Kadalmani Balamuthu at Mahatma Gandhi-Doerenkamp Center for Alternatives at Bharathidasan University.

Further, successful completion of my PhD study would not have been achieved unless the technical assistance of the followers Mr. Hasitha Weerathunga Doctoral research fellow at Industrial Training Institute for his generous help in GC-MS analysis, Dr. Thelma Jayasinghe for their guidance in NMR analysis, Dr. Chinthaka generous help for SEM image, Gowdhami Balakrishnan for training me in cell toxicity and molecular techniques, Mr. Sisira Kumara at Medical Research Institute and Dr. Tilusha Manchanayaka from Veterinary Institute Gannoruwa for animal toxicity studies. Prof. Piyal Marasinghe and Mr Gayan Kularathna from Nature Secret Pvt Ltd., also sincerely acknowledged for the support given in authentication of the plant. My appreciation also extended to, Mr. PAW Perera and Lalith sagara from Department of Chemistry the Open University of Sri Lanka for their support given during early in my research. Further, I would like to thank Hansani Karunarathne my laboratory colleague who shared the knowledge during the research.

Above all I would like to thank my beloved husband, Aruna and children, Savinuand Kenuka who have been very supportive, understanding and their patience. I wish to convey my gratitude to my beloved father and brother for their lifelongmoral support and encouragement. I dedicate this Thesis to my late mother.

## Reduction of cross contamination and extension of shelf life of ready to cook chicken meat treated with extracts of selected medicinal plants

#### Thikshani Somarathna

#### **ABSTRACT**

The aim of this study was to investigate the antimicrobial activity of less-utilized plants against food-borne pathogens *Escherichia coli*, *Salmonella*Typhimurium, *Listeria monocytogenes*, and *Staphylococcus aureus*, and to identify possible toxicity limits for safe use for extending the shelf life of ready to cook chicken meat.

In this study, four underutilized plants including *Alpinia malaccensis* ("Ran- kihiria"), *Terminalia catappa* ("Kottamba"), *Alpinia purpurata* ("Niyapothu mal") and *Curcuarma zedoaria* ("Haran-kaha") were extracted using ethanol or hexane solvents. The hexane rhizome extract of *A. malaccensis* showed significantly (P<0.05) higher diameter of inhibition zone ranging from 33.0±1.41 to 40.3±0.42 mm against *S. aureus* compared to other extracts. The lowest Minimal Inhibition Concentration (MIC) and Minimal Bactericidal Concentration (MBC) against *S. aureus* which was 20 and 80 mg/ml, respectively. GC-MS analysis showed an 82.87 % a major chemical compound for *A. malaccensis* whereas for *T. catappa* it was 2, 5-Furandione, 3- methyl (32.23%). The purified active fraction of *A. malaccensis* was identified by TLC bio-autography and it was confirmed as 1'Acetoxychavicol Acetate (1' ACA) by Nuclear Mangetic Resonance.

The combined plant extract of *A. malaccensis* (2.5 mg/ml) and *T. catappa* (20 mg/ml) had significant (P<0.05) synergistic antibacterial activity against *S. aureus* and *L. monocytogenes*. Also observed significantantibiofilm activity of *A. malaccensis* for both of cultures. The most effective combination was 5 mg/ml of *A. malaccensis* with 20 mg/ml of *T. catappa* for the antibiofilm activity.

A. malaccensis and T. catappa were used to evaluate the acute oral toxicity using Wistar rats and cytotoxicity study using cell culture techniques AO/EB, Hoechst staining and comet assay for A549, HepG2, 3T3 and COS7 cell lines. The acute oral toxicity studies for bothextracts at single dose of2000 mg/ kg body weightdid not cause any lethality or produce any remarkable changes in general behavior, body weights, biochemical parameters and histopathological studies. In addition, cytotoxicity results showed that non toxic concentration of A.malaccensis (2, 1.4, 30 and 1.4 μg/ml) and T. catappa (300, 300 and 130 μg/ml) were not mediated apoptotic cell death or necrosis, or DNA damage. Calculated approximately Annual Daily Intake for A. malaccensis and T. catappa were55.41g/day and 1549.70 mg/day. Therefore, the therapeutic levels for antibacterial activity of the plant extracts could be safe for consumption.

Finally, the use of both plant extracts in controlling *L.monocytogenes*, *S.aureus* and spoilage bacteria in vacuum packed ready-to-cook (RTC) marinated chicken was evaluated. Combination of both plant extracts significantly (p<0.05) inhibited the growth of *S. aureus* with 1.80, 2.13, 2.36 and 2.97 log cfu/g reduction over 3, 6, 9 and 12 days at 8 °C. Similarly, *L. monocytogenes* was significantly (p<0.05) inhibited at 6, 9, 12 days with 1.22, 1.60 and 1.55 log cfu/g reduction compared to control at 4 °C. Both temperatures significantly reduced (P<0.05) lipid oxidation in treated chicken compared to control. The shelf life of RTC marinated vacuum packed fresh chicken samples stored at 4 and 8 °C were significantly extended for 6 and 9 day respectively. Therefore, both plants extracts combination could use as antimicrobials and antioxidants in extending shelf life of RTC vaccum packed fresh chicken meat products.

Key words: Underutilized plant, Foodborne pathogens, A. malaccensis, 1'ACA, Chicken